Hypothesis

Apolipoprotein ɛ4-Associated Protection Against Pediatric Enteric Infections Is a Survival Advantage in Pre-Industrial Populations

Carr J. Smith^{a,*} and J. Wesson Ashford^b

^aSociety for Brain Mapping and Therapeutics, Pacific Palisades, CA, USA ^bStanford University and VA Palo Alto Health Care System, Palo Alto, CA, USA

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Abstract. Until 300,000 years ago, ancestors of modern humans ubiquitously carried the apolipoprotein E (*APOE*) ε 4/ ε 4 genotype, when the ε 3 allele mutated from the ancestral ε 4, which elevates the risk of Alzheimer's disease. Modern humans living today predominantly carry the ε 3 allele, which provides protection against heart disease and dementia in long-lived populations. The ancestral ε 4 allele has been highly preserved in isolated populations in tropical and Arctic regions with high pathogen burdens, e.g., helminths. Early humans experienced serious enteric infections that exerted evolutionary selection pressure, and factors that mitigate infant and childhood mortality from enteric infections also exert selection pressure. Some bacteria can exploit the host's defensive inflammatory response to colonize and invade the host. Pathogen-induced inflammation associated with infant and childhood diarrhea can damage the gut wall long after the invading organisms are no longer present. Inflammation not only resides in the mucosal wall, but also induces systemic inflammation. Baseline systemic inflammation is lower in ε 4 carriers, yet ε 4 carriers display a stronger host inflammatory response that reduces pathogen burdens, increasing infant and early childhood survival. Evolutionary selection of the ε 3 allele likely occurred after humans moved into temperate zones with lower pathogen burdens, unrelated to protection from Alzheimer's disease.

Keywords: Alzheimer's disease, ɛ4 allele, enteric infections, evolution, immunity, selection

THE APOLIPOPROTEIN ε3 ALLELE EVOLUTIONARILY OUTCOMPETED THE ε4 ALLELE

There are three common alleles of the apolipoprotein (APOE) gene, i.e., APOE $\varepsilon 2$, APOE $\varepsilon 3$, and APOE $\varepsilon 4$ [1]. In the general United States (US) population, the $\varepsilon 4$ allele prevalence is approximately 25.5%, constituted by a small portion of ε 4 homozygous individuals (2% of the US population) with a larger proportion being ε 3/ ε 4 (22%) and ε 2/ ε 4 heterozygous (1.5%) [2]. Until 300,000 years ago, ancestors of modern humans were ubiquitously ε 4/ ε 4, and then the ε 3 allele mutated from the ancestral ε 4 allele [3]. The ε 3 allele displayed a competitive survival advantage sufficiently robust to result in the current predominance of the *APOE* ε 3 gene, with a presence in approximately 96% of the US population with more than 60% being homozygous ε 3/ ε 3. One thought was that the ε 3 was successful because of its protection against atherosclerotic

^{*}Correspondence to: Carr J. Smith, PhD, DABT, 6400 Brindlewood Court, Mobile, AL 36608, USA. Tel.: +1 919 599 4517; E-mail: carrjsmith@yahoo.com.

heart disease, memory loss, and dementia in progressively older age ranges [4]. Similarly, the $\varepsilon 2$ allele mutated from the ε 3 allele about 200,000 years ago, but this protective allele has remained relatively rare, with the homozygous $\varepsilon 2/\varepsilon 2$ variant less than 1%, and $\varepsilon 2/\varepsilon 3$ heterozygosity occurring in about 11% of the population. The failure of the $\varepsilon 2$ allele to compete successfully against the $\varepsilon 3$ allele is not surprising given that its benefit on longevity and cognition [5], and protection from AD pathology [6, 7] does not contribute to survivability until an advanced age well after the average life expectancy, particularly of hunter gatherer populations. In addition, possession of the ε^2 allele is associated with elevated triglyceride levels, which can increase heart disease risk in middle-aged individuals, and predispose to certain infections that can occur in younger age groups [8, 9].

Possession of one $\varepsilon 4$ allele increases the risk of developing AD by 3 to 4-fold, and possession of two $\varepsilon 4$ alleles increases risk by 15-fold, as compared with the $\varepsilon 3/\varepsilon 3$ genotype, with a large part of the variation being related to substantially earlier age of onset. Over 60% of patients with "non-familial" AD carry the $\varepsilon 4$ allele [4, 10, 11]. This profound difference in AD risk is a consequence of conformational changes resulting from minor differences in the sequences of the *APOE* alleles. The three isoforms of *APOE* differ in amino acid sequence at only amino acid positions 112 and 158: the *APOE* $\varepsilon 2$ allele has cysteine at both positions and the *APOE* $\varepsilon 3$ allele has cysteine at 112 and arginine at 158 [12, 13].

ε4 ALLELE PREVALENCE IS HIGHER IN AREAS WITH GREATER PATHOGEN BURDENS

The prevalence of the ancestral apolipoprotein $\varepsilon 4$ allele is highest in hunter-gatherer populations residing in the high pathogen environments of the tropics or Arctic zones [14]. Tropical populations with a high prevalence of $\varepsilon 4$ carriers include the San people in Southern Africa [15], pigmies in the Congo [16], sub-Saharan tribes including the Fon, Zairians, and Tutsi [16], New Guineans [17], Malaysian and Australian aborigines [18], and the Sami of northern Scandinavia. In addition, the population frequency of the $\varepsilon 4$ allele decreases and the $\varepsilon 3$ allele increases when moving from southern to northern latitudes [18–21], until reaching the sub-Arctic and Arctic zones where the $\varepsilon 4$ allele begins to increase in frequency again [14].

Each of the ancient populations noted above experiences a significant burden of infectious disease including infestation from a variety of parasites endemic to the tropics or equatorial zones [22-24], or helminths found in the Arctic zone [25-32]. In contrast with other ancient peoples who live in hot weather climates, the Sami are descended from nomadic peoples who had inhabited northern Scandinavia for thousands of years [33]. The case of the ancient Sami people suggests that it is the parasite burden rather than the climate itself or latitude that is the critical factor in the persistence of the $\varepsilon 4$ allele. The Sami people have a very high ɛ4 frequency of 0.31, a level equivalent to that seen in current descendants of ancient tropical populations [34]. From several thousand to a few hundred years ago, the Sami hunted reindeer (domesticated caribou) and kept them in small numbers as pack and decoy animals [33]. For only a comparatively short time of their residency across northern Scandinavia, probably the past few hundred years, the Sami transitioned from nomadic hunting to reindeer herding.

The diet of the ancient nomadic Sami people can be extrapolated from dietary histories provided by current Sami peoples, some of whom are still nomadic. The ancestral Sami probably relied upon five main sources of protein: all edible parts of the reindeer including the blood; all edible parts of the moose; a cold freshwater fish related to salmon and trout called the arctic char; a freshwater fish related to salmon called the whitefish; and Kalix roe, i.e., the eggs of the saltwater vendace fish native to Bothian Bay on the coast between Sweden and Finland. Of these five protein sources, reindeer were and are the most important by far [35]. Throughout their circumpolar distribution, reindeer are known to be infected by several parasites including Brucellosis, foot rot, warble and bot flies, and gastrointestinal roundworms. Reindeer are frequently infected with Brucella suis biotype 4. Humans can become infected by eating or handling uncooked infected meat, milk, etc. The ancient Sami people would have been exposed to Brucellosis and other infectious organisms [36].

INFANT AND CHILDHOOD MORTALITY IS VERY HIGH IN HUNTER-GATHERERS

The San people of Southern Africa are the most ancient extant human population identified and studied [37–40]. The San people of this region had a population of several million until about 1,500 years ago when competition from cattle herders began pressuring the population downward [41] to its current levels of approximately 100,000 [42]. Twenty percent of San infants die within their first year, most commonly from gastrointestinal infections. Fifty percent of San children die before the age of 15 [41]. Also, adult San have a short average lifespan of only 45–50 years, with only 10% of the San population surviving past age 60 [41].

The high infant mortality rate in the San population is observed in other hunter-gatherer populations. Overall, infant mortality is over 30 times greater among hunter-gatherers than in the United States [43]. Early childhood mortality is over 100 times greater in hunter-gatherer populations than in the United States. On the average, only 57% of children survive to age 15 years in hunter-gatherer populations [43]. If a hunter-gatherer child reaches the age of 15, there is a 64% chance that he or she will live to or past the age of 45 [43].

ENTERIC INFECTIONS IN ε4 CARRIERS

Oria et al. [44] measured the degree of Giardia infections in stool samples in Brazilian children living in shantytowns. Carriers of the ε 4 allele had fewer Giardia infections [44]. The high bacterial count in the normal human gut results in an overall load of approximately one gram of endotoxin. The vast majority of this one gram does not access the bloodstream as only 100 ng of endotoxin injected intravenously induces inflammatory activation of the blood, somatic organs, and brain. As compared with *APOE* ε 3 carriers, *APOE* ε 4 carriers produce a stronger innate immune response when challenged by lipopolysaccharide (LPS). This result has been duplicated in *APOE* ε 4 mice [45].

In humans, vitamin D deficiency has been associated with increased risk of respiratory infections, *Clostridium difficile* infection of the GI tract, and bacterial sepsis. Ryz et al. (2015) [46] employed a mouse model to study the mechanism of vitamin D3 on susceptibility to enteric infection with *Citrobacter rodentium* (*C. rodentium*). This enteropathogenic bacteria can cause a self-limiting colitis in mice. Vitamin D3 deficiency in mice increased intestinal inflammation that killed some of the normal gut flora thereby allowing *C. rodentium* to overgrow and cause colitis. If the mice receive a relatively high dose of vitamin D3, the action of Th17 T-cells can be inhibited. In this murine model, too little or too much vitamin D3 can promote gut inflammation and increase susceptibility to enteric infection via *C. rodentium*. Huebbe et al. [47] demonstrated higher 25(OH)D (vitamin D) levels in *APOE* ε 4 targeted replacement mice as compared with *APOE* ε 3 and ε 2 mice. A similar allele association in elevation in vitamin D levels was observed in a small group of human research subjects (N = 93; *p* < 0.072), and in a larger general population sample (N = 699; *p* < 0.003).

Toll-like receptors (TLRs) are membranespanning receptors expressed on macrophages and dendritic cells. TLRs recognize specific proteins derived from bacteria. Gale et al. [48] showed that whole blood from healthy $\varepsilon 3/\varepsilon 4$ carriers stimulated by TLR2, TLR4, and TLR5 ligands induced more pro-inflammatory cytokine secretion than whole blood from $\varepsilon 3/\varepsilon 3$ carriers. In addition, following intravenous injection of LPS, $\varepsilon 3/\varepsilon 4$ carriers experienced higher hyperthermia and plasma TNF α , and earlier IL-6 release than $\varepsilon 3/\varepsilon 3$ carriers. Taken together, these results suggest a more robust innate immune response in humans who possess at least one copy of the $\varepsilon 4$ allele.

In a murine model that produced mice homozygous for either APOE ε_2 , ε_3 , or ε_4 , allele status was associated with significant differences in the composition of the gut biome. The relative abundance of Erysipelotrichia (bacteria of the phylum Bacillota) was associated with APOE ɛ4 status, a finding also seen in humans [49]. In a similar murine model, APOE ε 4 status was associated with a relative abundance of bacteria from the Lactobacillaceae family. This finding might be of clinical significance as an increase in Lactobacillus has been associated with localized acidification via lactic acid and hydrogen peroxide secretion. The acidic microenvironment is thought to reduce colonization by pathogenic bacteria including Cryptosporidium, and fungal infections [50].

APOE affects inflammatory processes in murine models of infection concomitant with malnutrition. Azevedo et al. [51] examined the role of *APOE* alleles in genetic strains of mice developed to model the effects of gut inflammation in a malnourished host. Mice were fed a low protein diet and infected with *Cryptosporidium parvum* (*C. parvum*). Transgenic *APOE* ε 4/ ε 4 mice had less weight loss than either transgenic ε 3/ ε 3 or wild-type mice, showed an accelerated pace of bacterial killing, and displayed better intestinal villi. *APOE* ε 4 contributes to a regulated cytokine response in *C. parvum* infected, transgenic $\varepsilon 4/\varepsilon 4$ mice characterized by increased ileal L-arginine selective cationic protein transporter (CAT-1), arginase-1 (possible marker of ileal recovery), and TLR9 transcripts. TLR9 binds bacterial and viral DNA thereby eliciting signaling cascades leading to release of pro-inflammatory cytokines. These data suggest that the $\varepsilon 4$ allele resists infection with *C. parvum* and assists in intestinal normalization postinfection.

The normal function of the inflammatory arm of the innate immune system is to combat infection and tissue injury [52]. Pathogens use many different defense mechanisms to aid in their invasion of their hosts [53], one of which is the hijacking of host inflammation to facilitate invasion and colonization [54]. The first demonstration of this facet of pathogen defense was reported in 1966 by Gray and Killenger [55]. They showed that Listeria monocytogenes invades macrophages recruited to the site of infection and travels inside the macrophages to distant tissues. Shigella recruit neutrophils across the epithelium, rupturing the epithelial barrier, thereby trading off some degree of neutrophilic killing of the Shigella for enhanced entry of the surviving bacteria into the host [56]. The pro-inflammatory cytokines interferon- γ and TNF- α can disrupt tight junctions between cells thereby increasing intercellular permeability [57].

Helicobacter pylori (H. pylori) uses gastric epithelial inflammation to establish long-term colonization [58]. Although H. pylori elicits a strong inflammatory response, the bacteria dampen host immune responses to facilitate chronic colonization of the stomach. CD4+ helper T cells are the most important T cell subtype involved in responses to H. pylori infection, with contributions from Th1 and Th17 cells, and regulatory T cells. H. pylori predominately elicits the production of Th1-type cytokines induced by IL-12 release by neutrophils and monocytes. H. pylori releases virulence factors that inhibit antigen presentation to T cells, block T cell proliferation, and suppress Th1 responses. Therefore, although H. pylori raises a strong innate immune response with attendant chronic inflammation, its immunosuppressive abilities facilitate bacterial persistence and long-term residence in the gastric mucosa [59].

Salmonella typhimurium (S. typhimurium) can invade the GI tract and cause diarrhea and acute intestinal inflammation eliciting a neutrophilic infiltrate. These bacteria enter and survive in mucosal macrophages residing in intestinal epithelium. In the inflamed gut, the growth of *S. typhimurium* is enhanced as other microbes are outcompeted. The mechanism underlying *Salmonella* overgrowth in intestinal inflammation is unclear. A possible mechanism is the reaction of inflammation-associated activated oxygen species with thiosulfate naturally resident in the gut to form tetrathionate, a respiratory electron acceptor. Tetrathionate allows *S. typhimurium* to utilize respiration and outcompete other gut microbes reliant on anerobic fermentation [60].

Several lines of evidence suggest that some pathogens use host inflammation to kill competitor pathogens. Brown et al. [25] have modeled the growth kinetics of how a particular pathogen can trigger host inflammation to the detriment of competitor pathogens. In humans, S. typhimurium induces a self-limiting gastroenteritis. Due to the similarity between human and porcine S. typhimurium-induced gastroenteritis, Chirullo et al. [61] used piglets to model Salmonellosis. These authors showed that wild-type strain S. typhimurium (STM14028) can employ host inflammation to enhance an active infection. STM14028 efficiently colonizes in vitro porcine monocytes-macrophages and intestinal columnar epithelial (IPEC-J2) cells. If the cells are pretreated with pro-inflammatory LPS, the degree of STM14028 colonization increases. If an LPS antagonist is used to inhibit the pro-inflammatory effect of LPS, the increased colonization is inhibited. In the in vivo arm of the Chirullo et al. [68] experiment, the pathogenicity of STM14028 in the presence and absence of inflammation was examined. When pro-inflammatory LPS is co-administered with S. typhimurium via the intraperitoneal (i.p.) route to piglets, at four hours post-infection both body temperature and blood cytokine levels are increased, consistent with the induction of acute inflammation. As compared with i.p. injection with S. typhimurium alone, co-administration of the bacteria with LPS showed a significant increase in colonization of tonsils, cecum, and spleen [61]. Also, Salmonella might derive nutrients from the inflammatory mucosa to stimulate growth. In addition, a gut wall highly colonized with different strains of bacteria represents a colonization barrier to new bacterial strain infection [62, 63]. Inflammation can kill a significant number of the bacterial strains colonizing the gut wall thereby facilitating the entry of a new infective bacterial strain [64]. Antibiotic treatment can illustrate this principle when normal gut flora are killed thereby facilitating

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the growth of more harmful strains, e.g. *Clostridium difficile* [65].

DAMAGE FROM PATHOGEN-INDUCED INFLAMMATION TO PEDIATRIC GUT STRUCTURE AND FUNCTION

Damage to the small bowel associated with pediatric diarrhea has a complex etiology [66]. Overgrowth of the normal microbiome of the gut by anaerobic bacteria such as Veillonella and Bacteroides species, predisposes to intestinal mucosal injury [67]. These and other strains of anaerobic bacteria chemically modify the primary bile acids cholic and chenodeoxycholic acid and convert them into deoxycholic and lithocholic acid, which are highly damaging to the jejunal mucosa. In the bowel lumen, deoxycholic and lithocholic acid induce water and sodium glucose malabsorption, increase intestinal permeability, and facilitate entry of intact and therefore potentially allergenic macromolecules. Dietary fats are solubilized in mixed micelles, which are disrupted by deoxycholic and lithocholic acid, leading to abnormal fecal excretion of dietary fats [66]. In rural West African children, enteropathy from chronic diarrhea has been shown to be mediated by T cells [68]. The pathological increase in intestinal permeability associated with chronic pediatric diarrhea has also been shown to cause endotoxemia, wherein LPS from intestinal bacteria enter the bloodstream. The presence of endotoxin (LPS) in the bloodstream leads to systemic inflammation [69]. While diarrhea initiates the enteropathy, post-initiation the enteropathy only co-exists with infant diarrhea for 10% of the time [70]. Once initiated, the enteropathy appears to perpetuate itself presumably via antigens passing from the intestinal lumen and into the mucosa where T-cell stimulation elicits further damage to the tissue [68].

ENTERIC INFECTIONS ARE A MAJOR CAUSE OF MORTALITY AND MORBIDITY IN HUNTER-GATHERER INFANTS AND CHILDREN

In surviving hunter-gatherer and extant preindustrial populations, enteric infections represent a major cause of mortality and morbidity in infants and children. For example, gastrointestinal infections are responsible for the majority of the 20% infant mortality rate seen in San People [41]. Although data on infant mortality due to enteric infection in ancestral hunter-gatherers are not available, today one in nine child deaths worldwide are due to diarrheal disease [71], the vast majority of which occur in less developed countries. Enteric infections are not only endemic in tropical and warm climates but are also endemic in hunter gatherer populations in the Arctic [72]. In 1959, Gordon and Babbott [73] reviewed the causes and frequency of acute intestinal infection in the Arctic. In Alaska, parasites were found in 62 of 227 residents in the Kuskokwim delta. In 1958, Fournelle et al. [74] examined 1,680 Alaskan Inuits and reported an infection rate of 40% for protozoa and 12% for helminths. Most of the helminths observed by Fournelle et al. [74] were diphyllobothriasis tapeworms found in raw or undercooked fish. Enteric infection rates were very high in Greenland, with 78 per cent of 274 Inuits examined having parasites with protozoa infection being about six times more common than helminth infection [73].

Early humans experienced enteric infections that have exerted a significant degree of evolutionary selection pressure [22]. Fumagalli et al. [22] calculated that the evolutionary pressure imposed by parasitic worms (helminths) on human genes has been stronger than pressure due to viral, protozoa, or bacterial agents. Based on results from Dunne and Cooke [75], Fumagalli et al. [22] have postulated that helminths evolve slower than unicellular/viral agents and have complex life cycles thereby resulting in relatively stable geographic distributions. Further, helminths and humans evolved on similar timescales that facilitated co-evolutionary interaction between the human host and parasite. Faster evolving species like viruses might not exert the same, or as long-lasting, selective pressure as helminths toward inducing change in ε 4 allele frequencies [22].

In 1961, HJ Heinz published a study on parasitical infections in ancient San people adult males who at that time were still living in the Kalahari Desert according to their ancestral lifestyle [76]. In 220 stool samples studied, the only pathogen found was the hookworm *Necator americanus*, as the many different species of parasitical worms that afflict other sub-Saharan African tribes are not able to survive in the arid conditions of the Kalahari [76]. Hookworm infections are more persistent than many other pathogens because of their ability to co-exist with the host by inhibiting the immune system. The Kalahari Desert is relatively wet from November to March. A San tribesman infected during this wet period during which hookworms can live outside of a host can continue to carry the hookworm infection into the dry season from April to October.

Loukas and Prociv [77] have reviewed immune responses to hookworms. In their review, these authors noted that protective immunity to hookworms does not develop in humans so that prolonged infections occur in all age groups. Both the larval stage and adult hookworms elicit a type 2 T-helper type immune response characterized by activated mast cells in gut mucosa, elevated serum IgE levels and eosinophils [77]. The lack of protective immunity against hookworm is consistent with the dry season infection rate of 26%, and wet season hookworm infection rate of 89.5% (17 out of 19 San men) observed by Heinz [76]. High hookworm infection rates in San adults implies that infants and children also experienced significant infection rates as infection is acquired by invasion of the infective larval stages through the skin [78].

Children and adults can display differential immune responses to the same enteric pathogen. *Shigella* is a bacterium that can cause enteritis that is usually more serious in children than in adults. Raqib et al. [79] showed that children with shigellosis have persistent activation of the innate immune response in the post-acute infection convalescent phase. The persistence of immune activation indicates delayed elimination of *Shigella* antigens in children [79].

APOE ε4 PROTECTS COGNITION IN PEDIATRIC ENTERIC INFECTIONS

Several large epidemiology studies have reported a relationship between elevations in systemic markers of inflammation and declines in cognition [80]. Inflammatory markers have suggested a proinflammatory state in cognitive decline including CD40 ligand, C-Reactive Protein (CRP), IL-6, soluble intracellular adhesion molecule-1, monocyte chemoattractant protein-1, myeloperoxidase, osteoprotegerin (OPG), P-selectin, TNF- α , and TNF receptor. Microglial cells are thought to play an important role in modulating neuroinflammatory processes related to the development of Alzheimer's disease [80].

Oria et al. [44, 81, 82] have shown that possession of the ε 4 allele protects against long-term cognitive deficits associated with severe diarrhea in Brazilian shanty-town children. The children were tested for verbal fluency according to the NEPSY Developmental Neuropsychological Assessment booklet. Coding test performance was evaluated using paired associated symbol recall (ages 8–16 years) in the standard WISC-111 protocol. To measure the child's ability to plan and to inhibit impulsive responses the children were administered Elithorn mazes (ages 8–16 years). The cognitive protection might be related to a greater availability of cholesterol to the developing brain in ε 4 carriers. In glial cells, *APOE* ε 4 knockout mice display an increased activity of the arginine cationic transporter thereby increasing nitric oxide levels. It can be postulated that the enterocytes lining the intestines of *APOE* ε 4 positive humans might also experience elevated nitric oxide levels which could enhance intestinal immunity [82].

Escherichia coli is the bacterial pathogen most commonly associated with endemic forms of childhood diarrhea [83], although several other pathogens are also common including Norovirus GII, shiga toxin-producing Escherichia coli (STEC), Giardia, and Sapovirus [24]. Trumble et al. [84] studied Amazonian Amerindians with high parasite burdens and found that the $\varepsilon 4$ allele is associated with improved cognitive function. In human populations not stratified by APOE allelic type, parasite infections have been shown to impair cognitive function [85-87]. Eosinophil counts rise in response to parasitic infections [88]. In homozygous $\varepsilon 3/\varepsilon 3$ carriers, higher eosinophil counts were associated with poor performance on all cognitive tests [84]. Adults who possessed at least one $\varepsilon 4$ allele ($\varepsilon 3/\varepsilon 4$ or $\varepsilon 4/\varepsilon 4$) and having high eosinophil counts indicating high levels of parasite infection demonstrated superior cognitive performance as compared with non-carriers. Despite similar environmental exposure to parasites, e4 carriers had significantly lower eosinophil counts than did non-carriers.

The Tsimane are an indigenous tribe of the lowland Bolivian Amazon. Despite high levels of infection and inflammation, they do not tend to develop arterial aging and cardiovascular disease. A clinical evaluation of 983 Tsimané women reported that 70% were infected with the parasitic roundworm *Ascaris lumbricoides*, which is believed to increase fertility by suppressing the immune system [89]. In a previous study on the Tsimane [90], serum CRP levels were 60% higher in $\varepsilon 3/\varepsilon 3$ carriers than in $\varepsilon 4$ carriers. These authors suggested that the elevated serum CRP levels indicate a higher parasite burden in the $\varepsilon 3$ homozygotes [90]. Figure 1 summarizes some of the complex interactions among possession of the $\varepsilon 4$ allele and the immune system.



Fig. 1. Interaction of $\varepsilon 4$ allele with the immune system.

SYSTEMIC INFLAMMATION IN POPULATIONS NOT INFECTED BY PARASITES IS RELATED TO APOE GENOTYPE (ε 4 CARRIERS < ε 3 CARRIERS < ε 2 CARRIERS)

CRP is a ring-shaped (annular) protein synthesized in the liver and composed of five subunits (pentameric). CRP plasma concentration increases in response to inflammation [91]. CRP production is stimulated by IL-6 [92]. Hubacek et al. [93] analyzed data from a large population sample of randomly selected individuals from seven Czech towns. Their analysis included data on 2,886 males and 3,344 females from the HAPIEE (Health, Alcohol, and Psychosocial factors In Eastern Europe) study. In both males and females, the lowest levels of plasma high sensitivity CRP were measured in the carriers of the $\varepsilon 4/\varepsilon 4$ and $\varepsilon 4/\varepsilon 3$ genotypes. As expected, two-thirds of the study subjects were $\varepsilon 3/\varepsilon 3$, i.e., the most common genotype. In this group, the median (interquartile range, IQR) concentration of high sensitivity CRP was 1.13 mg/l (IQR, 0.56–2.33) in men and 1.23 mg/l (IQR, 0.61–2.65) in women. In carriers of the $\varepsilon 4$ allele, the high sensitivity CRP level in males was 0.72 mg/l (IQR, 0.61–0.86) and 0.72 mg/l (IQR, 0.61–0.85) in females. The differences between the $\varepsilon 4$ non-carrier and $\varepsilon 4$ carrier groups were statistically significant (p < 0.001). Adjustment for age, sex, history of cardiovascular disease, or cardiovascular risk factors did not affect the result.

Kahri et al. [94] have also reported a statistically significant inverse relationship between possession of the ε 4 allele and plasma high sensitivity CRP lev-

els. This group measured the concentrations of serum high sensitivity CRP, soluble vascular cell adhesion molecule-1 (sVCAM-1), soluble intercellular adhesion molecule-1 (sICAM-1), and soluble E-selectin in 211 subjects from Finnish families with low high density lipoprotein (HDL) and in 157 normolipidemic subjects. Subjects carrying the ε 4 allele in both the low-HDL subgroup (p < 0.05) and the normolipidemic subgroup (p < 0.01) had lower concentrations of serum high sensitivity CRP than subjects without the ε 4 allele.

By conducting a large Korean study Yun et al. [95] validated that the inverse association between the $\varepsilon 4$ allele and CRP was not limited to European populations. This group evaluated the association of the APOE genotype with serum CRP levels and white blood cell count in two large Korean populationbased studies. The subjects analyzed were enrolled in the Dong-gu study and the Namwon Study. In the Dong-gu Study, 8,893 (3,525 men and 5,368 women) subjects \geq aged 50 years from the Dong-gu district of Gwangju Metropolitan City in South Korea were evaluated. In the Namwon Study, 10,032 (3,909 men and 6,123 women) subjects from the Namwon city of Jeollabuk do province in South Korea were analyzed. The polymerase chain reaction-restriction fragment length polymorphism method was used to identify APOE genotypes. After adjusting for age, sex, body mass index, smoking, diabetes, hypertension, and serum lipids, multivariable linear regression analysis was performed to evaluate the relationships among APOE genotypes and CRP levels and white blood cell count. White blood cell count did not differ among APOE genotypes. In the Dong-gu Study, ɛ4 heterozygotes ($\varepsilon 3/\varepsilon 4$) and $\varepsilon 4$ homozygotes ($\varepsilon 4/\varepsilon 4$) had significantly lower CRP levels as compared with the $\varepsilon 3/\varepsilon 3$ genotype, i.e., 0.50 mg/L versus 0.67 mg/L for the heterozygotes and 0.37 mg/L versus 0.67 mg/L for the homozygotes. In the Namwon Study, the $\varepsilon 4$ heterozygotes and the ε 4 homozygotes also had significantly lower CRP levels as compared with the $\varepsilon 3/\varepsilon 3$ genotype, i.e., 0.47 mg/L versus 0.66 mg/L for the heterozygotes and 0.45 mg/L versus 0.66 mg/L for the homozygotes.

CONCLUSIONS

The kinetics of the transition from the ubiquitous ancestral $\varepsilon 4/\varepsilon 4$ allelic distribution to the current 25.5% (23.5%% heterozygous; 2% homozygous) $\varepsilon 4$ prevalence in primarily European derived populations is unknown [2]. There is consensus that humans first evolved in Africa, with recent evidence supporting the coincidental evolution of geographically subdivided populations across Africa [96, 97]. Modern humans diverged from pre-human hominins 260,000 to 350,000 years ago [98]. The most recent evidence suggests that the vast majority of modern humans living today are descended from a population that migrated from southern Africa to eastern Africa about 70,000 years ago, and then traveled beyond the continent from East Africa approximately 60,000 years ago [39]. At the time humans migrated from southern Africa to eastern Africa, the area was hot and humid as it is today [99].

A strong host innate immune response capable of killing bacteria soon after infection is advantageous for infant and early childhood survival via reduction in pathogen burden and associated immune responses. Despite lower baseline levels of inflammation, ɛ4 carriers display enhanced innate immune responses [48]. The robust innate immune response seen in extant pre-industrial populations provides a parsimonious explanation for the persistence of the ε 4 allele (Fig. 2). A significant body of evidence suggests that an important normal function of the ε 4 allele was to reduce the morbidity and mortality from enteric infections in infants and children among hunter-gatherer populations. From this perspective, the ancestral ɛ4 allele is not defective as compared with the now more common $\varepsilon 3$ allele but is rather an adaptation to mankind's ancestral high-pathogen environment, still experienced by selected tropical and arctic modern human populations.

Concomitantly, a major question is what is the factor explaining the success of the $\varepsilon 3$ allele related to migration to temperate climates with a lower pathogen burden? While the advantage of the $\varepsilon 3$ allele for lowering cardiovascular disease and AD in elderly individuals may be significant [4], this possibility needs further anthropological study, as additional factors might have also contributed to the rapid success of the ɛ3 allele [100], including unexplained burdens of the $\varepsilon 4$ allele. The weight-of-the-evidence suggests that the $\varepsilon 4$ allele represents a case of antagonistic pleiotropy wherein natural selection originally favored the allele because it increased fitness and fertility early in life, although $\varepsilon 4$ increases the risk of AD later in life [4, 11, 101]. However, this primal allele may have yet other disadvantages in temperate climates and more recent human evolution. The more robust systemic inflammatory response in £4 carriers is consistent with the observation that neuroinflamAncestral human populations comprised of hunter-gatherers.

 \downarrow Infant and child mortality ~ 50% from enteric infections.

Enteric infections exerted strong evolutionary pressure.

↓

ε4 strongly selected for because of robust innate immunity and brain protection in infants with diarrhea.

↓

Humans began moving from southern Africa into lower-pathogen burden temperate zones.

1

Selection pressure for $\varepsilon 3$ allele increases.

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Modern humans in high-pathogen tropics and arctic retain a significant minority of $\varepsilon 4$ alleles while those in temperate zones are predominantly $\varepsilon 3$.

Fig. 2. Persistence of the $\varepsilon 4$ allele in high-pathogen environments.

mation might play a significant role in initiation or exacerbation of AD pathology. Further research on the possible relationship between systemic innate immunity and immune/inflammatory responses in the brain is warranted.

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REFERENCES

- [1] Yassine HN, Braskie MN, Mack WJ, Castor KJ, Fonteh AN, Schneider LS, Harrington MG, Chui HC (2017) Association of docosahexaenoic acid supplementation with Alzheimer disease stage in apolipoprotein E epsilon4 carriers: A review. JAMA Neurol 74, 339-347.
- [2] Evans DA, Beckett LA, Field TS, Feng L, Albert MS, Bennett DA, Tycko B, Mayeux R (1997) Apolipoprotein E epsilon4 and incidence of Alzheimer disease in

a community population of older persons. JAMA 277, 822-824.

- [3] Fullerton SM, Clark AG, Weiss KM, Nickerson DA, Taylor SL, Stengârd JH, Salomaa V, Vartiainen E, Perola M, Boerwinkle E, Sing CF (2000) Apolipoprotein E variation at the sequence haplotype level: Implications for the origin and maintenance of a major human polymorphism. *Am J Hum Genet* 67, 881-900.
- [4] Ashford JW (2004) APOE genotype effects on Alzheimer's disease onset and epidemiology. J Mol Neurosci 23, 157-165.
- [5] Sinclair LI, Pleydell-Pearce CW, Day INM (2017) Possible positive effect of the APOE ε2 allele on cognition in early to mid-adult life. *Neurobiol Learn Mem* 146, 37-46.
- [6] Serrano-Pozo A, Qian J, Monsell SE, Betensky RA, Hyman BT (2015) APOEepsilon2 is associated with milder clinical and pathological Alzheimer disease. *Ann Neurol* 77, 917-929.
- [7] Sinclair LI, Pleydell-Pearce CW, Day INM (2017) Possible positive effect of the APOE ε2 allele on cognition in early to mid-adult life. *Neurobiol Learn Mem* 146, 37-46.
- [8] Genest J (2003) Lipoprotein disorders and cardiovascular risk. *J Inherit Metab Dis* **26**, 267-287.
- [9] Freitas RS, Roque CR, Matos GA, Belayev L, de Azevedo OGR, Alvarez-Leite JI, Guerrant RL, Oriá RB (2022) Immunoinflammatory role of apolipoprotein E4 in malnutrition and enteric infections and the increased risk for chronic diseases under adverse environments. *Nutr Rev* 80, 1001-1012.
- [10] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261, 921-923.
- [11] Raber J, Huang Y, Ashford JW (2004) ApoE genotype accounts for the vast majority of AD risk and AD pathology. *Neurobiol Aging* 25, 641-650.

- [12] Mahley RW, Weisgraber KH, Huang Y (2009) Apolipoprotein E: Structure determines function, from atherosclerosis to Alzheimer's disease to AIDS. J Lipid Res 50(Suppl), S183-S188.
- [13] Huang Y, Mahley RW (2014) Apolipoprotein E: Structure and function in lipid metabolism, neurobiology, and Alzheimer's diseases. *Neurobiol Dis* 72(Pt A), 3-12.
- [14] Belloy ME, Napolioni V, Greicius MD (2019) A quarter century of APOE and Alzheimer's disease: Progress to date and the path forward. *Neuron* **101**, 820-838.
- [15] Sandholzer C, Delport R, Vermaak H, Utermann G (1995) High frequency of the apo allele in Khoi San from South Africa. *Hum Genet* 95, 46-48.
- [16] Zekraoui L, Lagarde JP, Raisonnier A, Gérard N, Aouizérate A, Lucotte G (1997) High frequency of the apolipoprotein E *4 allele in African pygmies and most of the African populations in sub-Saharan Africa. *Hum Biol* 69, 575-581.
- [17] Kamboh MI, Weiss KM, Ferrell RE (1991) Genetic studies of human apolipoproteins. ApoE polymorphism and cholesterol levels in the Mayans of the Yucatan peninsula, Mexico. *Clin Genet* **39**, 26-32.
- [18] Corbo RM, Scacchi R (1999) Apolipoprotein E (APOE) allele distribution in the world. Is APOE*4 a 'thrifty' allele? Ann Hum Genet 63, 301-310.
- [19] Gerdes LU (2003) The common polymorphism of apolipoprotein E: Geographical aspects and new pathophysiological relations. *Clin Chem Lab Med* **41**, 628-631.
- [20] Ewbank DC (2004) The APOE gene and differences in life expectancy in Europe. J Gerontol A 59, 16-20.
- [21] Hu P, Qin YH, Jing CX, Lu L, Hu B, Du PF (2011) Does the geographical gradient of ApoE4 allele exist in China? A systemic comparison among multiple Chinese populations. *Mol Biol Rep* 38, 489-494.
- [22] Fumagalli M, Sironi M, Pozzoli U, Ferrer-Admetlla A, Pattini L, Nielsen R (2011) Correction: Signatures of environmental genetic adaptation pinpoint pathogens as the main selective pressure through human evolution. *PLoS Genet* 7, e1002355.
- [23] Loy DE, Liu W, Li Y, Learn GH, Plenderleith LJ, Sundararaman SA, Sharp PM, Hahn BH (2017) Out of Africa: Origins and evolution of the human malaria parasites Plasmodium falciparum and Plasmodium vivax. *Int J Parasitol* 47, 87-97.
- [24] Lima AAM, Oliveira DB, Quetz JS, Havt A, Prata MMG, Lima IFN, Soares AM, Filho JQ, Lima NL, Medeiros PHQS, Santos AKS, Veras HN, Gondim RNDG, Pankov RC, Bona MD, Rodrigues FAP, Moreira RA, Moreira ACOM, Bertolini M, Bertolini LR, Freitas VJF, Houpt ER, Guerrant RL (2019) Etiology and severity of diarrheal diseases in infants at the semiarid region of Brazil: A case-control study. *PLoS Negl Trop Dis* 13, e0007154.
- [25] Brown SP, Le Chat L, Taddei F (2008) Evolution of virulence: Triggering host inflammation allows invading pathogens to exclude competitors. *Ecol Lett* 11, 44-51.
- [26] Arh I (1960) Fish tapeworm in Eskimos in the Port Harrison area, Canada. *Canad J Public Health* 51, 268-271.
- [27] Babbott FL Jr, Frye WW, Gordon JE (1961) Intestinal parasites of man in Arctic Greenland. Am J Trop Med Hyg 10, 185-190.
- [28] Laird M, Meerovitch E (1961) Parasites from northern Canada. I. Entozoa of Chime Eskimos. *Canad J Zool* 39, 63-67.

- [29] Cameron TWM, Choquette LPE (1963) Parasitological problems in high northern latitudes, with particular reference to Canada. *Polar Record* 11, 567-77.
- [30] Rausch RL, Scott EM, Rausch VR (1967) Helminths in Eskimos in western Alaska, with particular reference to Diphyllobothrium infection and anaemia. *Trans Roy Soc Trop Med Hyg* 61, 351-357.
- [31] Eaton RDP (1968) Amebiasis in northern Saskatchewan: Epidemiological considerations. *Canad Med Assoc J* 99, 706-711.
- [32] Freeman RS, Jamieson J (1976) Parasites of Eskimos at Igloolik and Hall Beach, Northwest Territories. In *Proceedings of the 3rd International Symposium* on Circumpolar Health, Yellowknife, NWT, Shephard RJ, Itoh S, eds. University of Toronto Press, Toronto, pp. 306-315.
- [33] Augustyn A. The Editors of Encyclopaedia Britannica (2019) Sami. https://www.britannica.com/topic/Sami, Date published January 9, 2020. Access date November 23, 2020.
- [34] Ross AB, Johansson A, Ingman M, Gyllensten U (2006) Lifestyle, genetics, and disease in Sami. *Croat Med J* 47, 553-565.
- [35] Veen, Ellis. Sami Food: What to eat in Lapland. September 2022, https://www.backpackadventures.org/sami-foodin-lapland/
- [36] Håglin L (1991) Nutrient intake among Saami people today compared with an old, traditional Saami diet. Arctic Med Res (Suppl), 741-746.
- [37] Schlebusch CM, Skoglund P, Sjödin P, Gattepaille LM, Hernandez D, Jay F, Li S, De Jongh M, Singleton A, Blum MG, Soodyall H, Jakobsson M (2012) Genomic variation in seven Khoe-San groups reveals adaptation and complex African history. *Science* 338, 374-379.
- [38] Veeramah KR, Wegmann D, Woerner A, Mendez FL, Watkins JC, Destro-Bisol G, Soodyall H, Louie L, Hammer MF (2012) An early divergence of KhoeSan ancestors from those of other modern humans is supported by an ABC-based analysis of autosomal resequencing data. *Mol Biol Evol* 29, 617-630.
- [39] Rito T, Richards MB, Fernandes V, Alshamali F, Cerny V, Pereira L, Soares P (2013) The first modern human dispersals across Africa. *PLoS One* 8, e80031.
- [40] Skoglund P, Thompson JC, Prendergast ME, Mittnik A, Sirak K, Hajdinjak M, Salie T, Rohland N, Mallick S, Peltzer A, Heinze A, Olalde I, Ferry M, Harney E, Michel M, Stewardson K, Cerezo-Román JI, Chiumia C, Crowther A, Gomani-Chindebvu E, Gidna AO, Grillo KM, Helenius IT, Hellenthal G, Helm R, Horton M, López S, Mabulla AZP, Parkington J, Shipton C, Thomas MG, Tibesasa R, Welling M, Hayes VM, Kennett DJ, Ramesar R, Meyer M, Pääbo S, Patterson N, Morris AG, Boivin N, Pinhasi R, Krause J, Reich D (2017) Reconstructing prehistoric African population structure. *Cell* **171**, 59-71.e21.
- [41] Kalahari Meerkat Project, Kalahari Bushmen, http://www.kalahari-meerkats.com, Date accessed November 23, 2020.
- [42] Kim HL, Ratan A, Perry GH, Montenegro A, Miller W, Schuster SC (2014) Khoisan hunter-gatherers have been the largest population throughout most of modern-human demographic history. *Nat Commun* 5, 5692.
- [43] Gurven M, Kaplan H (2007) Longevity among huntergatherers: A cross-cultural examination. *Popul Dev Rev* 33, 321-365.

- [44] Oriá RB, Patrick PD, Zhang H, Lorntz B, de Castro Costa CM, Brito GA, Barrett LJ, Lima AA, Guerrant RL (2005) APOE4 protects the cognitive development in children with heavy diarrhea burdens in Northeast Brazil. *Pediatr Res* 57, 310-316.
- [45] Brown GC (2019) The endotoxin hypothesis of neurodegeneration. J Neuroinflammation 16, 180.
- [46] Ryz NR, Lochner A, Bhullar K, Ma C, Huang T, Bhinder G, Bosman E, Wu X, Innis SM, Jacobson K, Vallance BA (2015) Dietary vitamin D3 deficiency alters intestinal mucosal defense and increases susceptibility to Citrobacter rodentium-induced colitis. *Am J Physiol Gastrointest Liver Physiol* **309**, G730-G742.
- [47] Huebbe P, Nebel A, Siegert S, Moehring J, Boesch-Saadatmandi C, Most E, Palla J, Egert S, Müller MJ, Schreiber S, Nöthlings U, Rimbach G (2011) APOE ε4 is associated with higher vitamin D levels in targeted replacement mice and humans. *FASEB J* 25, 3262-3270.
- [48] Gale SC, Gao L, Mikacenic C, Coyle SM, Rafaels N, Murray Dudenkov T, Madenspacher JH, Draper DW, Ge W, Aloor JJ, Azzam KM, Lai L, Blackshear PJ, Calvano SE, Barnes KC, Lowry SF, Corbett S, Wurfel MM, Fessler MB (2014) APOe4 is associated with enhanced *in vivo* innate immune responses in human subjects. *J Allergy Clin Immunol* 134, 127-134.
- [49] Zajac DJ, Green SJ, Johnson LA, Estus S (2022) APOE genetics influence murine gut microbiome. *Sci Rep* 12, 1906.
- [50] Parikh IJ, Estus JL, Zajac DJ, Malik M, Maldonado Weng J, Tai LM, Chlipala GE, LaDu MJ, Green SJ, Estus S (2020) Murine gut microbiome association with APOE alleles. *Front Immunol* 11, 200.
- [51] Azevedo OG, Bolick DT, Roche JK, Pinkerton RF, Lima AA, Vitek MP, Warren CA, Oriá RB, Guerrant RL (2014) Apolipoprotein E plays a key role against cryptosporidial infection in transgenic undernourished mice. *PLoS One* 9, e89562.
- [52] Newton K, Dixit VM (2012) Signaling in innate immunity and inflammation. *Cold Spring Harb Perspect Biol* 4, a006049.
- [53] Bhavsar AP, Guttman JA, Finlay BB (2007) Manipulation of host-cell pathways by bacterial pathogens. *Nature* 449, 827-834.
- [54] Pédron T, Sansonetti P (2008) Commensals, bacterial pathogens and intestinal inflammation: An intriguing ménage à trois. *Cell Host Microbe* 3, 344-347.
- [55] Gray ML, Killinger AH (1966) Listeria monocytogenes and listeric infections. *Bacteriol Rev* 30, 309-382.
- [56] Perdomo OJ, Cavaillon JM, Huerre M, Ohayon H, Gounon P, Sansonetti PJ (1994) Acute inflammation causes epithelial invasion and mucosal destruction in experimental shigellosis. J Exp Med 180, 1307-1319.
- [57] Bruewer M, Luegering A, Kucharzik T, Parkos CA, Madara JL, Hopkins AM, Nusrat A (2003) Proinflammatory cytokines disrupt epithelial barrier function by apoptosis-independent mechanisms. *J Immunol* **171**, 6164-6172.
- [58] Mimuro H, Suzuki T, Nagai S, Rieder G, Suzuki M, Nagai T, Fujita Y, Nagamatsu K, Ishijima N, Koyasu S, Haas R, Sasakawa C (2007) Helicobacter pylori dampens gut epithelial self-renewal by inhibiting apoptosis, a bacterial strategy to enhance colonization of the stomach. *Cell Host Microbe* 2, 250-263.
- [59] FitzGerald R, Smith SM (2021) An overview of Helicobacter pylori infection. In *Helicobacter Pylori. Methods*

in Molecular Biology, Smith SM, eds. Humana, New York, NY, vol. 2283.

- [60] Winter SE, Thiennimitr P, Winter MG, Butler BP, Huseby DL, Crawford RW, Russell JM, Bevins CL, Adams LG, Tsolis RM, Roth JR, Bäumler AJ (2010) Gut inflammation provides a respiratory electron acceptor for Salmonella. *Nature* 467, 426-429.
- [61] Chirullo B, Pesciaroli M, Drumo R, Ruggeri J, Razzuoli E, Pistoia C, Petrucci P, Martinelli N, Cucco L, Moscati L, Amadori M, Magistrali CF, Alborali GL, Pasquali P (2015) Salmonella Typhimurium exploits inflammation to its own advantage in piglets. *Front Microbiol* 6, 985.
- [62] Barnich N, Carvalho FA, Glasser AL, Darcha C, Jantscheff P, Allez M, Peeters H, Bommelaer Oria RB, Patrick PD, Oria MO, Lorntz B, Thompson MR, Azevedo OG, Lobo RN, Pinkerton RF, Guerrant RL, Lima AA (2010) APOE polymorphisms and diarrheal outcomes in Brazilian shanty town children. *Braz J Med Biol Res* 43, 249-256.
- [63] Stecher B, Robbiani R, Walker AW, Westendorf AM, Barthel M, Kremer M, Chaffron S, Macpherson AJ, Buer J, Parkhill J, Dougan G, von Mering C, Hardt WD (2007) Salmonella enterica serovar typhimurium exploits inflammation to compete with the intestinal microbiota. *PLoS Biol* 5, 2177-2189.
- [64] Lupp C, Robertson ML, Wickham ME, Sekirov I, Champion OL, Gaynor EC, Finlay BB (2007) Host-mediated inflammation disrupts the intestinal microbiota and promotes the overgrowth of Enterobacteriaceae. *Cell Host Microbe* 2, 119-129.
- [65] Beaugerie L, Petit JC (2004) Microbial-gut interactions in health and disease. Antibiotic-associated diarrhea. *Best Pract Res Clin Gastroenterol* 18, 337-352.
- [66] Andrade JAB, Fagundes-Neto U (2011) Persistent diarrhea: Still an important challenge for the pediatrician. J Pediatr 87, 199-205.
- [67] de Boisseau D, Chaussain M, Badoual J, Raymond J, Dupont C (1996) Small-bowel bacterial overgrowth in children with chronic diarrhea, abdominal pain, or both. J Pediatr 128, 203-207.
- [68] Campbell DI, Murch SH, Elia M, Sullivan PB, Sanyang MS, Jobarteh B, Lunn PG (2003) Chronic T cell-mediated enteropathy in rural west African children: Relationship with nutritional status and small bowel function. *Pediatr Res* 54, 306-311.
- [69] Campbell DI, Elia M, Lunn PG (2003) Growth faltering in rural Gambian infants is associated with impaired small intestinal barrier function, leading to endotoxemia and systemic inflammation. J Nutr 133, 1332-1338.
- [70] Lunn PG (2000) The impact of infection and nutrition on gut function and growth in childhood. *Proc Nutr Soc* 59, 147-154.
- [71] Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases (NCEZID), Division of Foodborne, Waterborne, and Environmental Diseases at CDC (2020) Global diarrhea burden. https://www.cdc.gov/healthywater/global/ Date accessed November 23, 2020. Date last reviewed December 17, 2015.
- [72] Brown M, Green JE, Boag TJ, Kuitunen-Ekbaum E (1950) Parasitic infections in the Eskimos at Igloolik, N.W.T. *Canad J Public Health* **41**, 508-512.
- [73] Gordon JE, Babbott FL (1959) Acute intestinal disease in the Arctic. Am J Public Health Nations Health 49, 1441-1453.

- [74] Fournelle HJ, Wallace IL, Rader VA (1958) Bacteriological and parasitological survey of enteric infections in an Alaskan Eskimo area. *Am J Public Health Nations Health* 48(11 Pt 1), 1489-1497.
- [75] Dunne DW, Cooke A (2005) A worm's eye view of the immune system: Consequences for evolution of human autoimmune disease. *Nat Rev Immunol* 5, 420-426.
- [76] Heinz HJ (1961) Factors governing the survival of bushmen worm parasites in the Kalahari. S Afr J Sci 57, 207-213.
- [77] Loukas A, Prociv P (2001) Immune responses in hookworm infections. *Clin Microbiol Rev* 14, 689-703.
- [78] Hotez PJ, Bethony J, Bottazzi ME, Brooker S, Buss P (2005) Hookworm: "The great infection of mankind". *PLoS Med* 2, e67.
- [79] Raqib R, Moly PK, Sarker P, Qadri F, Alam NH, Mathan M, Andersson J (2003) Persistence of mucosal mast cells and eosinophils in shigella-infected children. *Infect Immun* 71, 2684-2692.
- [80] Gorelick PB (2010) Role of inflammation in cognitive impairment: Results of observational epidemiological studies and clinical trials. *Ann N Y Acad Sci* **1207**, 155-162.
- [81] Oriá RB, Patrick PD, Blackman JA, Lima AA, Guerrant RL (2007) Role of apolipoprotein E4 in protecting children against early childhood diarrhea outcomes and implications for later development. *Med Hypotheses* 68, 1099-107.
- [82] Oria RB, Patrick PD, Oria MO, Lorntz B, Thompson MR, Azevedo OG, Lobo RN, Pinkerton RF, Guerrant RL, Lima AA (2010) APOE polymorphisms and diarrheal outcomes in Brazilian shanty town children. *Braz J Med Biol Res* 43, 249-256.
- [83] Benevides-Matos N, Pieri FA, Penatti M, Orlandi PP (2015) Adherence and virulence genes of Escherichia coli from children diarrhea in the Brazilian Amazon. *Bras J Microbiol* 46, 131-137.
- [84] Trumble BC, Stieglitz J, Blackwell AD, Allayee H, Beheim B, Finch CE, Gurven M, Kaplan H (2017) Apolipoprotein E4 is associated with improved cognitive function in Amazonian forager-horticulturalists with a high parasite burden. *FASEB J* **31**, 1508-1515.
- [85] Katan M, Moon YP, Paik MC, Sacco RL, Wright CB, Elkind MS (2013) Infectious burden and cognitive function: The Northern Manhattan Study. *Neurology* 80, 1209-1215.
- [86] Gale SD, Erickson LD, Brown BL, Hedges DW (2015) Interaction between Helicobacter pylori and latent toxoplasmosis and demographic variables on cognitive function in young to middle-aged adults. *PLoS One* 10, e0116874.
- [87] Gale SD, Erickson LD, Berrett A, Brown BL, Hedges DW (2016) Infectious disease burden and cognitive function in young to middle-aged adults. *Brain Behav Immun* 52, 161-168.
- [88] O'Connell EM, Nutman TB (2015) Eosinophilia in infectious diseases. *Immunol Allergy Clin North Am* 35, 493-522.
- [89] Gurven M, Kaplan H, Winking J, Rodriguez DE, Vasunilashorn S, Kim JK, Finch C, Crimmins E (2009) Inflammation and infection do not promote arterial aging and cardiovascular disease risk factors among lean horticulturalists. *PLoS One* 4, e6590.

- [90] Vasunilashorn S, Finch CE, Crimmins EM, Vikman SA, Stieglitz J, Gurven M, Kaplan H, Allayee H (2011) Inflammatory gene variants in the Tsimane, an indigenous Bolivian population with a high infectious load. *Biode-mography Soc Biol* 57, 33-52.
- [91] Thompson D, Pepys MB, Wood SP (1999) The physiological structure of human C-reactive protein and its complex with phosphocholine. *Structure* 7, 169-177.
- [92] Del Giudice M, Gangestad SW (2018) Rethinking IL-6 and CRP: Why they are more than inflammatory biomarkers, and why it matters. *Brain Behav Immun* 70, 61-75.
- [93] Hubacek JA, Peasey A, Pikhart H, Stavek P, Kubinova R, Marmot M, Bobak M (2010) APOE polymorphism and its effect on plasma C-reactive protein levels in a large general population sample. *Hum Immunol* 71, 304-308.
- [94] Kahri J, Soro-Paavonen A, Ehnholm C, Taskinen MR (2006) ApoE polymorphism is associated with C-reactive protein in low-HDL family members and in normolipidemic subjects. *Mediators Inflamm* 2006, 12587.
- [95] Yun YW, Kweon SS, Choi JS, Rhee JA, Lee YH, Nam HS, Jeong SK, Park KS, Ryu SY, Choi SW, Kim HN, Cauley JA, Shin MH (2015) APOE polymorphism is associated with C-reactive protein levels but not with white blood cell count: Dong-gu Study and Namwon Study. J Korean Med Sci 30, 860-865.
- [96] Scerri EML, Thomas MG, Manica A, Gunz P, Stock JT, Stringer C, Grove M, Groucutt HS, Timmermann A, G. Rightmire P, d'Errico F, Tryon CA, Drake NA, Brooks AS, Dennell RW, Durbin R, Henn BM, Lee-Thorp J, de Menoca P, Petraglia MD, Thompson JC, Scally A, Chikhi L (2018) Did our species evolve in subdivided populations across Africa, and why does it matter? *Trends Ecol Evol* 33, 582-594.
- [97] Hershkovitz I, Weber GW, Quam R, Duval M, Grün R, Kinsley L, Ay A (2018) The earliest modern humans outside Africa. *Science* 359, 456-459.
- [98] Schlebusch CM, Malmström H, Günther T, Sjödin P, Coutinho A, Edlund H, Munters AR, Vicente M, Steyn M, Soodyall H, Lombard M, Jakobsson M (2017) Southern African ancient genomes estimate modern human divergence to 350,000 to 260,000 years ago. *Science* 358, 652-655.
- [99] Scholz CA, Johnson TC, Cohen AS, King JW, Peck JA, Overpeck JT, Talbot MR, Brown ET, Kalindekafe L, Amoako PYO, Lyons RP, Shanahan TM, Castañeda IS, Heil CW, Forman SL, McHargue LR, Beuning KR, Gomez J, Pierson (2007) East African megadroughts between 135 and 75 thousand years ago and bearing on early-modern human origins. *Proc Natl Acad Sci U S A* 104, J16416-J16421.
- [100] Theendakara V, Peters-Libeu CA, Bredesen DE, Rao RV (2018) Transcriptional effects of ApoE4: Relevance to Alzheimer's disease. *Mol Neurobiol* 55, 5243-5254.
- [101] Corbett S, Courtiol A, Lummaa V, Moorad J, Stearns S (2018) The transition to modernity and chronic disease: Mismatch and natural selection. *Nat Rev Genet* 19, 419-430.